# Synthesis, Transformations and Biological Activity of Chloro Enamines and Ynamines Derived from Chloroalkenyl- and Alkynyl-*N*-substituted Purine and Pyrimidine Bases of Nucleic Acids<sup>1</sup>

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Reactions of nucleic acid bases and related heterocycles 1-5, 18 and 22 with tetra-, tri- and dichloroethylenes 6-9 in hexamethylphosphoric triamide gave the corresponding N-trichloro-, -dichloro- and -chloro- enamines 11-17, 19, 20, 23 and 24 in high regioselectivity (Nº for purines and N<sup>1</sup> for pyrimidines). Bases 1, 5, 18, 22 and trichloroethylene 7 gave the respective E-dichloro enamines 15, 16, 20 and 24. Compounds 16 and 20 were identical with the products obtained by addition of bases 1 and 18 to dichloroacetylene. Thymine 18 and compound 7 gave  $N^1, N^3$ -bisdichloro enamine 21 as the major product. The latter exists at room temperature as a mixture of rotamers **28** and **29** ( $\Delta G^{\ddagger} \cong 18$  kcal mol<sup>-1</sup>). The reaction of adenine **1** with (Z)-1,2- or 1,1dichloroethylene 8 or 9 furnished Z-chloro enamine 17 whereas thymine 18 and tetrachloroethylene 6 in dimethyl sulfoxide afforded a reduction product 20. Benzoylation of N<sup>9</sup>-(trichlorovinyl)adenine 11 gave  $N^6$ ,  $N^6$ -dibenzoyl derivative 26. The reaction of  $N^1$ -(dichlorovinyl) cytosine 24 with N, Ndimethylformamide dimethyl acetal afforded amidine 25. Interaction of (E)-N<sup>9</sup>-(dichlorovinyl)adenine 16 with sodium methoxide gave exclusively E-enamine 27. Trichloro enamines 11-14, 19, **23** and **26** were transformed to ynamines **30–35**. Hydrogenation of compounds **30** and **35** furnished  $N^9$ -ethyladenine **36** and  $N^1$ -ethylthymine **37**. Alkylation of ynamine **30** with acetone **38** gave only carbinol 41 whereas cyclohexanone 39 gave both compound 42 and cyclic ketal 43. The reaction of ynamines 30 and 35 with ketone 40 afforded only ketals 44 and 45. The reaction of compound 30 with N.N-dimethylformamide dimethyl acetal led to N-dimethylaminomethylene derivative 46. Ynamine 30 is a substrate for adenosine deaminase.

Heterocyclic ynamines containing a terminal acetylene moiety have not been investigated in any detail.<sup>2</sup> Very recently, a single compound of this class, *N*-ethynylpyrrole, was described <sup>3</sup> but it was characterized only by <sup>1</sup>H NMR spectroscopy. In connection with our studies of unsaturated nucleoside analogues<sup>4</sup> we became interested in ynamines derived from nucleic acid bases. Such compounds could serve as synthetic intermediates for the preparation of new analogues with a therapeutic potential but they could also exhibit some interesting biological properties *per se*. In addition, they may also become a source of the corresponding polymers (polyacetylenes)<sup>5</sup> containing nucleic acid bases. Similar polymeric materials, polyvinyl analogues of nucleic acids, inhibit RNA polymerase and exhibit other biological effects.<sup>6</sup>

In this communication, we describe the synthesis of chloro enamines 11–17, 19, 20, 23–27, ynamines 30–35, and some of their physicochemical and biological properties, as well as alkylation of compounds 30 and 35 with selected ketones. The rotational isomerism of bis-dichloro enamine 21 (rotamers 28 and 29) will also be discussed.

Synthesis of Chloro Enamines.—The chloro enamines were considered as convenient intermediates for the synthesis of ynamines derived from nucleic acid bases. The base-catalysed reaction of purines 1–5 (Scheme 1) and pyrimidines 18 and 22 (Schemes 2 and 3) with chlorinated ethylenes 6–9 was selected as a method of choice for synthesis of chloro enamines 11–17, 19, 20, 23 and 24. In the case of dichlorovinyl derivatives 15, 16, 20 and 24, this approach avoided use of the explosive and toxic dichloroacetylene used for the synthesis of (E)-N-(1,2-dichlorovinyl)imidazole.<sup>7</sup> A similar method was employed in the preparation of *N*-ethynyl derivatives of secondary amines<sup>8,9</sup> and *N*-ethynylpyrrole.<sup>3</sup>

Selection of a suitable solvent was crucial for successful reaction with chlorinated ethylenes 6–9. The appropriate heterocyclic bases 1–5, 18 and 22 were converted into the sodium or potassium salts by using NaH or KH, but only poor yields or side-reactions were observed in dimethylformamide (DMF) or dimethyl sulfoxide (DMSO). By contrast, hexamethylphosphoric triamide (HMPA) gave consistent though moderate yields of the desired intermediates 11–17, 19, 20, 23 and 24 (20–35%). In the case of reaction of adenine 1 with 1,1dichloroethylene 9, a phase-transfer method<sup>10</sup> employing NaOH and NBu<sub>4</sub>F (TBAF) in DMF gave better results than reaction in HMPA. 2-Amino-6-chloropurine 5 did not react with tetrachloroethylene 6 but trichloroethylene 7 gave compound 15 in 26% yield. The reaction of adenine 1 and (Z)-1,2-dichloroethylene 8 or 1,1-dichloroethylene 9 afforded the





Scheme 1 Reagents and conditions: i, NaH, HMPA; ii, KH, HMPA; iii, NaOH, NMe<sub>4</sub>F, DMF



Scheme 2 Reagents and conditions: i, NaH, HMPA; ii, NaH, DMSO



Scheme 3 Reagents and conditions: i, NaH, HMPA; ii, Me<sub>2</sub>NCH-(OMe)2, DMF

same product, (Z)-N-(2-chlorovinyl)adenine 17. The (E)-1,2dichloroethylene 10 did not react. It is of interest that the reaction of thymine 18 with tetrachloroethylene 6 led only to  $N^{1}$ -(trichlorovinyl)thymine 19 but trichloroethylene 7 gave bisdichloro enamine 21 in 28% yield and only a 6% yield of (E)- $N^1$ -(dichlorovinyl)thymine 20. This is, to the best of our knowledge, the first indication that trichloroethylene 7, a recognized nephrotoxic compound,11 mutagen and cancersuspect agent,<sup>12</sup> is capable of reacting with a functional group present in DNA and involved in Watson-Crick base-pairing. It is then clear that tetrachloroethylene 6 and trichloroethylene 7 differ in reactivity (see also compound 15 discussed above), possibly because of a change in the reaction mechanism (vide supra). The reaction of thymine 18 with NaH and tetrachloroethylene 6 in DMSO led to a reductive removal of one chlorine atom to give compound 20, identical with that obtained by reaction with trichloroethylene 7. No bis-dichloro enamine was detected.

All these reactions except that of compounds 18 and 7 proceeded with a high degree of regioselectivity with respect to the heterocyclic base. Thus,  $N^9$  is a preferential site of attack for purines and  $N^1$  for pyrimidines. In that respect, the selectivity of alkylation followed the pattern found for reactions of nucleic acid bases with alkylating agents having a leaving group attached to an sp<sup>3</sup>-hybridized carbon.<sup>13</sup> This was corroborated by the UV and <sup>13</sup>C NMR spectra. Thus, compounds 11, 16 and 17 exhibit  $\lambda_{max}$  256–260 nm similar to those of  $N^9$ -alkyladenines.<sup>13</sup> A similar pattern was found for purines 12, 13 and 15. The <sup>13</sup>C NMR spectrum of compound 13 (heterocyclic moiety) has a pattern similar to that found in  $N^9$ , 6-O-dialkylpurines.<sup>14</sup> The UV spectra of pyrimidines 19, 20, 23 and 24 are of lesser value for regioisomeric assignments. Thus, both thymine derivatives 19 and 20 have  $\lambda_{max}$  260 nm, which is similar to that of  $N^1, N^3$ -bis-dichloro enamine 21  $(\lambda_{max} 264 \text{ nm})$ . Similarly, the UV maximum usually associated with N<sup>1</sup>-alkylcytosines at ~ 275 nm<sup>13</sup> is apparently a part of the broad band at  $\lambda_{max}$  250 nm in compounds 23 and 24. Additional evidence for regioisomeric assignments in both purine and pyrimidine chloro enamines will be discussed later. The transformations described herein are stereoselective, giving only a single geometric isomer as seen from the <sup>1</sup>H and <sup>13</sup>C NMR spectra. It was shown that reactions of nucleophiles (enolates) with trichloroethylene 7 involved an intermediary formation of dichloroacetylene and, hence, a trans-stereochemistry.15 Therefore, a similar reaction course (eliminationaddition) was anticipated with chlorinated ethylenes 8 and 9. This is also compatible with the finding that, as mentioned earlier, adenine 1 and chloride 8 or 9 gave exclusively the Zisomer 17. The fact that (E)-1,2-dichloroethylene 10, which cannot form chloroacetylene by a trans-elimination, was unable to react with adenine 1 is in accord with such a mechanism.<sup>16</sup> It should be noted that N-(1,2-dichlorovinyl)pyrrole, prepared by reflux of the potassium salt of pyrrole in trichloroethylene 7, was formulated as the Z-isomer<sup>3</sup> but no evidence was offered to support this assignment.

These conclusions were further supported by reaction of dichloroacetylene with adenine 1 and thymine 18. As already mentioned, the former reagent and imidazole gave (E)-N-(1,2)dichlorovinyl)imidazole as a single product.<sup>7</sup> The reaction of adenine 1 with dichloroacetylene in HMPA was accompanied by extensive formation of coloured by-products and it gave dichloro enamine 16 in only 3% yield and 77% purity. Nevertheless, this compound was identical (TLC, mixed TLC, UV and <sup>1</sup>H NMR) with the product obtained from adenine 1 and trichloroethylene 7 as mentioned above. The reaction of thymine 18 with dichloroacetylene was much cleaner and afforded compound 20 in 12% yield, which was identical with a sample obtained from trichloroethylene 7. It is noteworthy that the latter reaction was significantly more regioselective than that of thymine 18 with trichloroethylene 7. Only traces of bisdichloro enamine 21 were detected in the reaction mixture.

It is obvious that an elimination-addition mechanism via the respective acetylenes is precluded in reactions with tetrachloroethylene 6 resulting in the formation of N-trichlorovinyl derivatives 11-14, 19 and 22. The reaction course most likely follows an addition-elimination pattern.<sup>16</sup>

The di- or tri-chlorovinyl moieties of 11 and 24 are stable enough to allow protection by benzoylation or reaction with N,N-dimethylformamide dimethyl acetal<sup>17</sup> (Schemes 3 and 4). The respective products 26 and 25 were obtained in 56 and 85% yield, respectively. The reaction of  $(E)-N^9-(1,2-\text{dichlorovinvl})$ adenine 16 with MeONa in DMF was highly regio- and stereoselective, and afforded a single product in 63% yield (Scheme 5). Since an elimination-addition mechanism can be excluded, four regio- and stereo-isomers, all resulting from an additionelimination mechanism, are possible. Absence of isomeric



Scheme 4 Reagents and conditions: i, PhCOCl, pyridine



Scheme 5 Reagents and conditions: i, MeONa, DMF

products complicated the structural assignment. Nevertheless, similar nucleophilic substitutions are known to proceed with stereoconvergence or retention of the original stereochemistry.<sup>18</sup> Such a reasoning would favour structure **27** for the product.

The 2D <sup>1</sup>H and <sup>13</sup>C NMR spectrum indicated that the heterocyclic proton signal located at higher field ( $\delta$  8.15) belongs to 8-H. Also, the signal at  $\delta_{\rm C}$  147.43 is the hydrogencarrying vinylic carbon C-2'. The C-1' signal is then found downfield, at  $\delta$  102.33, which is in accord with similar shifts observed in vinyl ethers.<sup>19</sup> The only significant nuclear Overhauser effect (NOE) was observed for the 2'-H and MeO (5.3 and 12%, respectively). This suggested a cis or geminal relationship for both functions. It should be noted that the NOE alone could not be used for confirmation of the regioand stereo-isomeric assignment because of an inherent ambiguity in distinguishing between isomeric olefins related by a 1,2-trans interchange of substituents.<sup>20</sup> Smaller NOEs (1-2.8%) were observed between 8-H and 2'-H in compound 27 and dichloro enamine 16. The distance between both hydrogens is 3.9 and 4.0 Å as determined for the energy-minimized (Chem3D Plus, Version 3.0) structures of compounds 16 and 27. Similar distances between relevant hydrogen atoms led to NOEs of comparable magnitude as noted for various conformationally restricted nucleosides.<sup>21</sup>

The electron-impact (EI) mass spectrum contains peaks assignable to fragments carrying adenine and a chlorine atom at the same carbon  $(m/z \ 184, \ 182; \ Scheme \ 6)$ . The latter peak is derived from the molecular ion by a loss of methyl group followed by hydrogen shift and decarbonylation. A similar set of ions originating from the adenine moiety of the respective molecular ions by a consecutive loss of one  $(m/z \ 155, \ 157)$  and two  $(m/z \ 128, \ 130)$  HCN units <sup>22</sup> was also present. This type of fragmentation is fully compatible with structure **27**.

The NMR spectra of  $(E,E)-N^1,N^3$ -bis(dichlorovinyl)thymine 21 deserve special comment. The 300 MHz <sup>1</sup>H NMR spectrum in CDCl<sub>3</sub> showed olefinic protons 2'- and 2"-H as singlets but doubling of some signals was observed in the 75 MHz <sup>13</sup>C NMR spectrum. The 500 MHz <sup>1</sup>H NMR spectrum in (CD<sub>3</sub>)<sub>2</sub>SO indicated that all proton peaks were doubled (Fig. 1) whereas the corresponding 125 Mz <sup>13</sup>C NMR spectrum contained 14 signals, indicating a splitting of all carbons except C-1', -2' (or -2"), -5 and Me (Fig. 2). Interestingly, in CDCl<sub>3</sub> at 500 MHz none of the proton signals was split but the comparable <sup>13</sup>C NMR spectrum exhibited 15 peaks (Fig. 3). Thus, the C-1' signal, which was observed in (CD<sub>3</sub>)<sub>2</sub>SO as a single peak, was



Scheme 6

split in two. Also, the pattern of C-2' and -2" signals was rearranged. These results can be readily explained by the presence of two rotational isomers 28 and 29. It is important to add that the 500 MHz <sup>1</sup>H and 125 MHz <sup>13</sup>C NMR spectra of (E)-N<sup>1</sup>-(dichlorovinyl)thymine 20 did not show any doubling of signals. The latter observation supports the assignment of structure 20 and, by analogy, trichloro enamine 19 as N<sup>1</sup>-regioisomers.



The final confirmation came from a temperature-dependence study of <sup>1</sup>H and <sup>13</sup>C NMR spectra (Figs. 1 and 2). Thus, all proton signals collapsed between 60-80 °C to the respective singlets or doublets. The free-activation energy  $\Delta G^{\ddagger}$  of the rotational isomerization is 17.7 kcal mol<sup>-1</sup> \* as calculated from the Eyring equation.<sup>24</sup> The latter value is in excellent agreement with the value  $\Delta G^{\ddagger}$  17.8 kcal mol<sup>-1</sup> obtained from the <sup>13</sup>C NMR spectrum. Although it was possible to determine the ratio of both rotamers as 3:2 from integrated signals of the <sup>1</sup>H NMR spectrum, we have been unable to assign the resonances to distinct rotameric forms. It should be noted that the two rotameric (anti and syn) forms of  $N^3$ -( $\beta$ -D-glucopyranosyl)-6methyluracil were discernible by <sup>1</sup>H NMR spectroscopy in the same ratio.<sup>25</sup> Similar rotamers were also noted in some  $\beta$ -D-glucopyranosylpteridines.<sup>26</sup> In these cases, the rotation of the sugar residue is hindered by two carbonyl groups or a carbonyl function and a heteroaromatic moiety. The obtained  $\Delta G^{\ddagger}$ -values,<sup>25,26</sup> 18.9 and 18.3 kcal mol<sup>-1</sup> (the former was calculated from the reported<sup>25</sup> data), indicate a similar rotational barrier in all these compounds. To the best of our knowledge, rotamers 28 and 29 are the first examples of such isomerism in a non-carbohydrate derivative of a nucleic acid base, which is also reflected in the <sup>13</sup>C NMR spectrum.

Synthesis and Properties of Ynamines.—The trichlorovinyl derivative 11 was smoothly converted into ynamine 30 in almost 60% yield by using BuLi in tetrahydrofuran (THF) at -70 °C (Scheme 7). A low temperature is essential for the success of

<sup>\*</sup> 1 cal = 4.184 J.

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Fig. 1 The temperature dependence of the 500 MHz <sup>1</sup>H NMR spectrum of rotamers 28 and 29 in  $(CD_3)_2$ SO. Only the olefinic region is shown. All values of chemical shifts at 23 °C are listed in the Experimental section. For numbering of signals see formula 21. Note that all protons of the less abundant rotamer are located downfield from those of the major rotational isomer. The spectra were determined at 23, 40, 60, 70 and 80 °C. At 80 °C all peaks were singlets or, where appropriate, doublets. The  $\Delta G^{\dagger}$ -value 17.7 kcal mol<sup>-1</sup> represents an average of determinations from the 2'- and 2"-H signals.

the reaction; at higher temperatures extensive decomposition occurred. Other chlorinated derivatives such as 16 and 17 did not react, probably because of limited solubility in THF at -70 °C. For these reasons, trichlorovinyl derivatives 12–14, 19, 23 and 26 were chosen for generation of the corresponding ynamines 31–35. The yields were in the range of ~40–50% with the exception of compound 32 which was obtained in only 21% yield. Dichlorovinyl derivative 15 which is soluble in THF did not afford any ynamine. This can be explained by preferential ionization of the C<sup>8</sup>–H bond of the heterocyclic moiety<sup>27</sup> which decreases the reactivity at the acetylenic CH function. Attempted debenzylations of compound 32 using boron tri-



**30**, **36**; B = adenin- $N^9$ -yl **31**; B = 2,6-diaminopurin- $N^9$ -yl **32**; B = 2-amino-6-benzyloxypurin- $N^9$ -yl **33**; B =  $N^6$ -benzoyladenin- $N^9$ -yl **34**; B = cytosin- $N^1$ -yl **35**, **37**; B = thymin- $N^1$ -yl **38**; R<sup>1</sup> = R<sup>2</sup> = Me **39**; R<sup>1</sup>R<sup>2</sup> = [CH<sub>2</sub>]<sub>5</sub> **40**; R<sup>1</sup> = R<sup>2</sup> = PhCH<sub>2</sub>OCH<sub>2</sub> **41**; B = adenin- $N^9$ -yl, R<sup>1</sup> = R<sup>2</sup> = Me **42**; **43**; B = adenin- $N^9$ -yl, R<sup>1</sup> = R<sup>2</sup> = PhCH<sub>2</sub>OCH<sub>2</sub> **44**; B = adenin- $N^9$ -yl, R<sup>1</sup> = R<sup>2</sup> = PhCH<sub>2</sub>OCH<sub>2</sub> **45**; B = thymin- $N^1$ -yl, R<sup>1</sup> = R<sup>2</sup> = PhCH<sub>2</sub>OCH<sub>2</sub>

Scheme 7 Reagents and conditions: i, BuLi, THF, --70 °C; ii, NaNH<sub>2</sub>, THF; iii, H<sub>2</sub>, Pd/C, EtOH



Fig. 2 The temperature dependence of the 125 MHz <sup>13</sup>C NMR spectrum of rotamers 28 and 29 in  $(CD_3)_2$ SO. See Fig. 1. Only the relevant segments of the spectrum are shown. Note that 5 carbons from a possible 9 are doubled. All were single peaks at 80 °C. The  $\Delta G^{\ddagger}$ -value 17.8 kcal mol<sup>-1</sup> is an average of determinations from the C-4, C-2, C-6 and C-1″ signals.



Fig. 3 The 125 MHz <sup>13</sup>C NMR spectrum of the olefinic region of rotamers 28 and 29 in CDCl<sub>3</sub>. See Fig. 1. Note that the signal of the C-1' carbon is split and the pattern of the C-2' and -2'' peaks is reversed relative to the spectrum determined in (CD<sub>3</sub>)<sub>2</sub>SO (Fig. 2). Assignments of the C-1', -1'' and -2' (-2'') signals are based on a DEPT experiment.<sup>23</sup>

chloride, trimethylsilyl iodide or sodium in liquid ammonia led only to side-reactions at the ynamine moiety.

The protection of ynamines is of interest in view of the considerable synthetic potential of such derivatives. The  $N^6$ -dimethylaminomethylene ynamine **46** was smoothly obtained

from compound **30** by reaction with N,N-dimethylformamide dimethyl acetal<sup>17</sup> in DMF in 75% yield (Scheme 8). By contrast,



Scheme 8 Reagents and conditions: i, Me<sub>2</sub>NCH(OMe)<sub>2</sub>, DMF

an attempted benzoylation of compound 30 with benzoyl chloride in pyridine led only to products of addition across the triple bond. Nevertheless, the  $N^6$ -benzoyl ynamine 33 was obtained from  $N^6, N^6$ -dibenzoyl derivative 26 (18%) or, more conveniently,  $N^6$ -benzamide 14 (47% yield) by a procedure outlined above. These transformations conclusively showed that reaction of  $N^6$ -benzoyladenine 4 with tetrachloroethylene 6 exhibits the same (N<sup>9</sup>) regioselectivity as that of adenine 1.

The obtained ynamines are of sufficient stability to be properly characterized.  $N^1$ -Ethynylcytosine 34 is an exception. Solutions of compound 34 in DMSO or DMF turn rapidly brown. The NMR spectra in  $(CD_3)_2NCDO$  at -20 °C revealed the presence of signals assignable to olefinic protons which are indicative of a spontaneous polymerization of compound 34. Hydrogenation of ynamines 30 and 35 afforded the corresponding N-ethyl derivatives 36 and 37 (Scheme 7) in 88 and 86% yield, respectively. The latter transformations also showed that reactions of adenine 1 and thymine 18 with tetrachloroethylene 6 were regioselective at N<sup>9</sup> and N<sup>1</sup>, respectively. Also, the UV spectrum of compound 34 is similar to that of cytallene,<sup>28</sup> which is in line with the formulation of trichloro enamine 23 as an N<sup>1</sup>-substituted cytosine.

The structures of the obtained ynamines were confirmed by spectroscopic methods. The presence of a C=CH function was apparent from IR and NMR spectra. The C-1' signals of pyrimidine ynamines are positioned downfield from those of purine derivatives. The trend appears to be the opposite for C-2'. The EI mass spectra of ynamines indicated little cleavage of the C-1'-N bond (peaks of heterocyclic bases are virtually absent). In this respect, the spectra are quite similar to those of other unsaturated derivatives of nucleic acid bases which contain a double bond in conjugation with the heterocyclic moiety.<sup>4</sup>

Reactions of Ynamines 30 and 35 with Ketones.—Alkylation of alkyl- or aryl-acetylenes with aldehydes or ketones is a general and synthetically useful transformation.<sup>29</sup> By contrast, reaction of ketones with ynamines derived from strong bases (e.g., diethylamine) leads only to rearrangement products of the intermediary carbinols. Less basic ynamines, such as N-ethynyl-N,N-diphenyl- or N-ethynyl-N-methyl-N-phenylamine, can be alkylated with ketones to the corresponding carbinols without difficulty.<sup>30</sup> No such hydroxyalkyl ynamines have been described in the heterocyclic series to the best of our knowledge. We therefore studied the reaction of ynamine 30 with acetone 38, cyclohexanone 39 and dibenzyloxyacetone  $^{31}$  40. The character of the base catalyst has a profound influence on the success of the alkylation. Thus, experiments using LiNH<sub>2</sub> (38) or the strongly basic BuLi (39 or 40) were fruitless. By contrast, reaction of ynamine 30 with acetone 38 in the presence of NaNH<sub>2</sub> in THF gave the corresponding carbinol 41 in 45% yield (Scheme 7). In a similar fashion, cyclohexanone 39 afforded carbinol 42 (70%) accompanied by cyclic unsaturated ketal 43 (29%). Carbinol 42 is an obvious intermediate in the formation of ketal 43. Thus, reaction of alcohol 42 with a second molecule of ketone 39 leads to the corresponding hemiketal<sup>1b</sup>

which then undergoes cyclization to the product ketal 43. Alkylation of ynamines 30 and 35 with ketone 40 afforded only cyclized products 44 and 45 in 72 and 54% yield, respectively. It should also be noted that carbinols 41 and 42 are stable during chromatography on silica gel and they thus differ from their counterparts derived from *N*-methyl- or *N*-phenyl-aniline.<sup>30</sup>

It is therefore clear that the reactivity of both the ketone (hemiketal formation) and the ynamine play a role in this transformation. Cyclization driven by formation of a hemiketal is generally not observed during alkylation of alkyl- or arylacetylenes with aldehydes or ketones but it was described in the case of reactive diacetylene carbinols<sup>32</sup> and formaldehyde. In accord with this trend, interaction of ynamine 30 with acetone 38 gave only carbinol 41, but reaction of cyclohexanone 39 with ynamine 30 furnished some cyclized product 43. Neither pyrimidine nor purine ynamines (30, 35) with ketone 40 afforded any carbinol, but gave only cyclic ketals (44 or 45). It has to be stressed that changing the reaction conditions, limiting the amount of ketone, and protecting the heterocyclic moiety (ynamines 33 and 46) did not influence the result. Ketals 44 and 45 were always the major products (after deprotection where appropriate) and the corresponding carbinols were not detected at all. A favourable effect of NaNH<sub>2</sub> in these transformations is difficult to explain. However, an interaction of NaNH<sub>2</sub> with the heterocyclic moiety, e.g. that of ynamine 30, may lead to transient formation of a base 47 (M = Na) stronger than the starting ynamine and, hence, increase the reactivity of the acetylene CH function. By contrast, BuLi will only decrease such reactivity by ionization of the amino group of ynamine 30. The less basic LiNH<sub>2</sub> is apparently not capable of providing intermediate 47 (M = Li).



Assignments of six-membered  $\beta$ -vinyl ether structures 43-45 followed from the NMR spectra. The <sup>13</sup>C NMR spectrum of compound 44 showed two olefinic carbons (C-6' and -5') at  $\delta_{\rm C}$  146.14 and 92.96, respectively. The spectrum of compound 45 contained similar signals at  $\delta_{\rm C}$  147.54 and 97.30. The signal at higher field was assigned to C-5' as in other  $\beta$ -vinyl ethers<sup>19</sup> (see also compound 27). In the case of compound 44, a distortionless enhancement by polarization transfer (DEPT) experiment<sup>23</sup> confirmed that C-5' was the hydrogen-carrying carbon. Likewise, the structures of carbinols 41 and 42 were unambiguously established by IR and NMR spectra which confirmed the presence of OH and C=C functions in both molecules. The UV spectra of carbinols 41 and 42 revealed some important differences in their fine structure (Fig. 4). Thus, compound 42 resembles the parent ynamine 30 more than does the dimethylcarbinol 41.

Biological Activity.—Compound 30 is a substrate of moderate activity for adenosine deaminase from calf intestine.<sup>1b</sup> Thus, ~90% deamination was observed after incubation for 3 days at room temperature.  $N^9$ -(prop-2-ynyl)adenine,<sup>33</sup> a homologue of compound 30,  $N^9$ -vinyladenine and ynamine 31 were inactive. Previous data <sup>1b</sup> indicated that compounds 19, 30, 31 and 42 inhibited the growth of murine leukaemia L1210 with IC<sub>50</sub> 40–150 µmol dm<sup>-3</sup> as determined by a clonogenic assay.<sup>13</sup> In addition, analogues 11, 30 and 31 suppressed the growth of mouse and human colon tumours C38, H8 or H116 according to a disk-diffusion assay.<sup>13</sup> Therefore, compounds 11 and 31



Fig. 4 UV spectra of ynamines 30, 41 and 42 in ethanol: ---- compound 30; ---- compound 41; ···· compound 42

were tested *in vivo* against early-stage colon adenocarcinoma C38 in mice. No antitumour activity was observed at a total dose of 1968 and 1100 mg kg<sup>-1</sup>, respectively.

Analogues 3, 33, 16 and 24 were also inactive as antiviral agents against herpes simplex virus (HSV-1) and human cytomegalovirus (HCMV) grown in culture  $^{34,35}$  at concentrations 100 µmol dm<sup>-3</sup> or lower. With the exception of compound 3, they were also not cytotoxic in human foreskin fibroblasts (HFF) or KB cells. Compound 3 inhibited the growth of KB cells with IC<sub>50</sub> ~ 100 µmol dm<sup>-3</sup>.

## Experimental

For general methods see refs. 28, 36. THF was freshly distilled from sodium benzophenone ketyl. DMF, DMSO, pyridine and HMPA were dried with molecular sieves 3 Å. Column chromatography and TLC were performed in the following solvent systems:  $S_1$ ,  $CH_2Cl_2$ -acetone (1:1);  $S_2$ ,  $CH_2Cl_2$ -MeOH (9:1);  $S_3$ ,  $CH_2Cl_2$ -MeOH (4:1);  $S_4$ ,  $CH_2Cl_2$ -MeOH (95:5);  $S_5$ , ethyl acetate-MeOH (95:5);  $S_6$ ,  $CH_2Cl_2$ -acetone (4:1);  $S_7$ ,  $CH_2Cl_2$ -acetone (9:1);  $S_8$ , hexane-acetone (7:3). Spots of samples containing high boiling solvents (HMPA) were dried under reduced pressure.<sup>37</sup> The UV and NMR spectra were determined in ethanol and ( $CD_3$ )<sub>2</sub>SO, respectively, unless stated otherwise. *J*-Values are given in Hz. For IR spectra, KBr pellets were used. For chemical ionization (CI) mass spectra, 2-methylpropane was used as the ionization gas.

 $N^9$ -(*Trichlorovinyl*)adenine 11. Adenine 1 (2 g, 15 mmol) was dissolved in HMPA (130 cm<sup>3</sup>) at 60 °C (bath temperature). The solution was cooled to room temperature and NaH (57% oil suspension; 1.24 g, 30 mmol) was added under N<sub>2</sub>. The mixture was stirred till the evolution of H<sub>2</sub> ceased. Tetrachloroethylene 6 (7.36 g, 44.5 mmol) was then added and the mixture was stirred at 60 °C (bath temp.) for 15 h. The progress of reaction was monitored by TLC in solvent S<sub>1</sub>. Most of the HMPA was removed under reduced pressure at 90 °C (bath temp.) and the resultant crude product was washed successively with methanol (20 cm<sup>3</sup>) and several times with portions (50 cm<sup>3</sup>) of solvent system S<sub>2</sub>. The combined washings were evaporated and the residue was flash chromatographed on a silica gel column, using solvent system S<sub>1</sub>. The appropriate fractions were combined and evaporated to give *compound* 11 (0.84 g, 20%), m.p. 189–190 °C (decomp.). This product was recrystallized from solvent system S<sub>2</sub> (0.1 g/50 cm<sup>3</sup>), m.p. 201–202 °C (decomp.) (Found: C, 31.8; H, 1.4; Cl, 40.4; N, 26.2. C<sub>7</sub>H<sub>4</sub>Cl<sub>3</sub>N<sub>5</sub> requires C, 31.79; H, 1.52; Cl, 40.21; N, 26.48%);  $\lambda_{max}/nm$  256 ( $\varepsilon$  15 000);  $\delta_{H}$  8.48 and 8.21 (2 H, 2 s, 2- and 8-H) and 7.58 (2 H, s, NH<sub>2</sub>);  $\delta_{C}$  156.53, 154.56, 149.56, 140.11 and 118.09 (adenine) and 125.27 and 120.59 (C-1', -2'); *m/z* (EI) 267, 265 and 263 (M, 5, 15 and 15%), 232, 230 and 228 (M - Cl, 11, 64 and 100) and 205, 203 and 201 (M - HCN - Cl, 2, 17 and 28) (Found: M<sup>+</sup>, 262.9528. C<sub>7</sub>H<sub>4</sub><sup>35</sup>Cl<sub>3</sub>N<sub>5</sub> requires M, 262.9532).

(E)-N<sup>9</sup>-(1,2-Dichlorovinyl)adenine 16.—A. From adenine 1 and trichloroethylene 7. The experiment was performed as described for compound 11 but with adenine 1 (0.27 g, 2 mmol), KH (24% oil suspension, 0.68 g, 4 mmol) and trichloroethylene 7 (0.26 g, 2 mmol) in HMPA (total 10 cm<sup>3</sup>). The reaction mixture was stirred for 16 h at room temperature. Work-up and chromatography (solvent system S<sub>2</sub>) followed similar procedures described for compound 11, to give title compound 16 (0.1 g, 22%), m.p. 229-230 °C after crystallization from solvent system S<sub>3</sub> (Found: C, 36.7; H, 2.4; Cl, 30.6; N, 30.6. C<sub>7</sub>H<sub>5</sub>Cl<sub>2</sub>N<sub>5</sub> requires C, 36.55; H, 2.19; Cl, 30.82; N, 30.44%);  $\lambda_{max}/nm$  258 ( $\epsilon$ 15 100);  $\delta_{\rm H}$  8.42 and 8.19 (2 H, 2 s, 2- and 8-H) and 7.52 (3 H, apparent s, NH<sub>2</sub> and 2'-H);  $\delta_{C}$  156.26, 153.81, 149.09, 139.41 and 117.89 (adenine) and 122.05 and 119.55 (C-1', -2'); m/z (EI) 233, 231 and 229 (M, 2, 10 and 16%), 196 and 194 (M - Cl, 32 and 100) and 169 and 167 (M - HCN - Cl, 7 and 21) (Found:  $M^+$ , 228.9919.  $C_7H_5{}^{35}Cl_2N_5$  requires M, 228.9922).

B. From adenine 1 and dichloroacetylene. Dichloroacetylenediethyl ether complex <sup>7</sup> (7.5 cm<sup>3</sup>, ~12 mmol) was added dropwise to a solution of adenine 1 (0.27 g, 2 mmol) in HMPA (10 cm<sup>3</sup>). The mixture, which gradually became orange, was stirred at room temperature for 1 h before being evaporated, and HMPA was removed at 70 °C (bath-temp.) and 0.025 mmHg pressure. The crude product was chromatographed on a silica gel column in solvent system S<sub>4</sub> to give dichloro enamine 16 (25 mg, 3%), m.p. 220–225 °C, identical (TLC, <sup>1</sup>H NMR) with a sample prepared by method A. Compound 16 was 77% pure as estimated by UV spectroscopy.

(Z)-N<sup>9</sup>-(2-Chlorovinyl)adenine 17.—A. From adenine 1 and (Z)-1,2-dichloroethylene 8. A mixture of adenine 1 (0.1 g, 0.74 mmol) and KH (0.25 g, 1.5 mmol) in HMPA (18 cm<sup>3</sup>) was stirred at room temperature for 2 h. Reagent 8 (0.14 cm<sup>3</sup>, 1.5 mmol) was added and the mixture was stirred at 65 °C (bath temp.) for 7 h. The usual work-up and chromatography in solvent system S<sub>2</sub> gave product 17 (50 mg, 35.4%), m.p. 218-219 °C (decomp.) after crystallization from ethanol (Found: C, 43.0; H, 3.0; Cl, 18.4; N, 35.65. C<sub>7</sub>H<sub>6</sub>ClN<sub>5</sub> requires C, 42.98; H, 3.09; Cl, 18.12; N, 35.80%;  $\lambda_{max}/nm$  260 ( $\varepsilon$  12 600);  $\delta_{H}$  8.62 and 8.16 (2 H, 2 s, 2- and 8-H), 7.44 (1 H, d, J<sub>1',2'</sub> 6.3, 1'-H), 7.41 (2 H, s, NH<sub>2</sub>) and 6.61 (1 H, d,  $J_{2,1}$  6.3);  $\delta_{C}$  156.49, 153.69, 149.37, 138.75 and 117.80 (adenine) and 121.52 and 111.19 (C-1', -2'); m/z (EI) 197 and 195 (M, 17 and 50%), 160 (M - Cl, 100), 133 (M - HCN - Cl, 44), 106 (M -  $2 \times$  HCN - Cl, 19) (Found: M<sup>+</sup>, 195.0308. C<sub>7</sub>H<sub>6</sub><sup>35</sup>ClN<sub>5</sub> requires M, 195.0312).

B. From adenine 1 and 1,1-dichloroethylene 9. Reagent 9 (0.72 cm<sup>3</sup>, 9 mmol) was added to a mixture of adenine 1 (0.2 g, 1.5 mmol), NaOH (0.24 g, 6 mmol), 1 mol dm<sup>-3</sup> TBAF in THF (0.2 cm<sup>3</sup>, 0.2 mmol) and 0.5 cm<sup>3</sup> water in DMF (20 cm<sup>3</sup>). The mixture was stirred at 60 °C (bath temp.) for 24 h and then was evaporated. Chromatography of the residue in solvent system S<sub>5</sub> gave compound 17 (39 mg, 16%), m.p. 220 °C

(decomp.) which was identical with the product obtained by method A.

2,6-Diamino-N<sup>9</sup>-(trichlorovinyl)purine 12.—The procedure described for compound 11 was followed with 2,6-diaminopurine hemisulfate 2 (2.19 g, 11 mmol), NaH (60%, 1.66 g, 42 mmol) and tetrachloroethylene 6 (5.45 g, 33 mmol) in HMPA (100 cm<sup>3</sup>) to give compound 12 (1.03 g, 33.1%), m.p. 196–199 °C after crystallization from solvent system S<sub>1</sub> (Found: C, 29.6; H, 2.1; Cl, 37.3; N, 29.4. C<sub>7</sub>H<sub>5</sub>Cl<sub>3</sub>N<sub>6</sub>-0.25 H<sub>2</sub>O requires C, 29.60; H, 1.95; Cl, 37.44; N, 29.59%);  $\lambda_{max}/nm$  278 ( $\varepsilon$  12 400);  $\delta_{H}$  7.95 (1 H, s, 8-H), 6.95 (2 H, s, 6-NH<sub>2</sub>) and 6.17 (2 H, s, 2-NH<sub>2</sub>);  $\delta_{c}$  161.57, 156.75, 152.05, 135.60 and 112.18 (purine) and 124.27 and 121.66 (C-1', -2'); *m/z* (EI) 284, 282, 280 and 278 (M, 0.1, 4, 13 and 13%), 247, 245 and 243 (M - Cl, 1, 10 and 15) and 205, 203 and 201 (M - HN=C=NH - Cl, 1, 8 and 13) and 49 (100).

2-Amino-6-benzyloxy-N<sup>9</sup>-(trichlorovinyl)purine 13.—2-Amino-6-benzyloxypurine <sup>38</sup> 3 (1.20 g, 5.3 mmol), NaH (60%, 0.43 g, 10.6 mmol) and tetrachloroethylene 6 (2.64 g, 16 mmol) in HMPA (100 cm<sup>3</sup>) were allowed to react at 60 °C for 15 h. The usual work-up and chromatography in solvent system S<sub>7</sub> afforded compound 13 (0.4 g, 20%), m.p. 171–172 °C after crystallization from hexane–acetone (4:1) (Found: C, 45.2; H, 2.9; Cl, 28.5; N, 18.7.  $C_{14}H_{10}Cl_3N_5O$  requires C, 45.37; H, 2.72; Cl, 28.70; N, 18.90%);  $\lambda_{max}/mm$  270 ( $\varepsilon$  13 200);  $\delta_H$  8.15 (1 H, s, 8-H), 7.47, 7.36 and 7.34 (5 H, d and m, Ph), 6.90 (2 H, s, NH<sub>2</sub>) and 5.46 (2 H, s, CH<sub>2</sub>);  $\delta_C$  161.14, 160.79, 154.22, 137.99 and 112.97 (purine), 136.68, 129.08, 128.84 and 128.59 (Ph), 121.89 and 121.09 (C-1', -2') and 67.68 (CH<sub>2</sub>); m/z (EI) 375, 373, 371 and 369 (M, 0.6, 6, 20 and 21%) and 91 (C<sub>6</sub>H<sub>5</sub>CH<sub>2</sub>, 100).

N<sup>6</sup>-Benzoyl-N<sup>9</sup>-(trichlorovinyl)adenine 14.—A mixture of N<sup>6</sup>-benzoyladenine 4 (0.3 g, 1.25 mmol), NaH (60%, 0.1 g, 2.5 mmol) and tetrachloroethylene 6 (0.62 g, 3.8 mmol) was stirred in HMPA (15 cm<sup>3</sup>) for 14 h at 60 °C (bath temp.). After the usual work-up, chromatography in solvent system S<sub>6</sub> gave compound 14 (0.1 g, 20%), m.p. 158–160 °C after crystallization from the same solvent mixture (Found: C, 45.65; H, 2.0; Cl, 28.7; N, 19.1. C<sub>14</sub>H<sub>8</sub>Cl<sub>3</sub>N<sub>5</sub>O requires C, 45.62; H, 2.19; Cl, 28.85; N, 19.00%);  $v_{max}$ /cm<sup>-1</sup> 3260–3220 (NH) and 1720 (CO);  $\lambda_{max}$ /nm 278 (ε 16 900);  $\delta_{\rm H}$  11.42 (CONH), 8.83 (2 H, s, 2-and 8-H), 8.04 (2 H, d) and 7.62–7.50 (3 H, m) (Ph); *m/z* (EI) 371, 369 and 367 (M, 1, 4 and 4%), 344, 342, 340 and 338 (M – 29, 1, 10, 30 and 30) and 105 (PhCO, 100).

(E)-2-Amino-6-chloro-N<sup>9</sup>-(1,2-dichlorovinyl)purine 15.—2-Amino-6-chloropurine 5 (0.1 g, 0.6 mmol), NaH (60%; 47 mg, 1.2 mmol) and trichloroethylene 7 (0.23 g, 1.7 mmol) were allowed to react in HMPA (10 cm<sup>3</sup>) for 15 h at room temperature. The crude product was chromatographed in solvent system S<sub>6</sub> to give compound 15 (40 mg, 26%), m.p. 195– 196 °C after crystallization from solvent system S<sub>2</sub> (Found: C, 31.9; H, 1.6; Cl, 40.4; N, 26.3. C<sub>7</sub>H<sub>4</sub>Cl<sub>3</sub>N<sub>5</sub> requires C, 31.79; H, 1.52; Cl, 40.21; N, 26.48%);  $\lambda_{max}/nm$  302 ( $\varepsilon$  6100) and 247sh ( $\varepsilon$  7700);  $\delta_{\rm H}$  7.90 (1 H, s, 8-H), 7.38 (1 H, s, 2'-H) and 6.15 (2 H, s, NH<sub>2</sub>);  $\delta_{\rm C}$  161.14, 153.85, 150.93, 141.56 and 122.86 (purine) and 122.16 and 121.15 (C-1', -2'); m/z (FAB) 269, 267 and 265 (M + H, 12, 32 and 34%) and 58 (100).

N<sup>1</sup>-(*Trichlorovinyl*)thymine 19.—The procedure for compound 11 was followed on a 16 mmol scale of thymine 18. After evaporation of HMPA the residue was chromatographed in solvent system S<sub>6</sub>. The appropriate fractions were evaporated to give, after trituration with solvent system S<sub>8</sub>, *solid* 19 (1 g, 25%), m.p. 189–191 °C (softens at 165–170 °C) (Found: C, 32.7; H, 2.05; Cl, 41.8; N, 11.0.  $C_7H_5Cl_3N_2O_2$  requires C, 32.91; H, 1.97; Cl, 41.63; N, 10.96%);  $\lambda_{max}/nm$  259 ( $\varepsilon$  10 700);  $\delta_{H}$ 11.84 (1 H, s, NH), 7.71 (1 H, d, 6-H) and 1.76 (3 H, apparent s, Me);  $\delta_{C}$  164.16, 148.64, 138.47, 112.66 and 12.43 (thymine), 125.77 and 125.45 (C-1', -2'); m/z (EI) 260, 258, 256 and 254 (M, 0.2, 2, 4 and 4%), 223, 221 and 219 (M – Cl, 11, 64 and 100), 217, 215, 213 and 211 (M – HCNO, 1, 10, 31 and 32), 180, 178 and 176 (M – HCNO – Cl, 2, 9 and 14), and 152, 150 and 148 (M – HCNO – CO – Cl, 9, 40 and 54).

(E)-N<sup>1</sup>-(1,2-Dichlorovinyl)thymine 20.—A. Reductive alkylation of thymine 18 with tetrachloroethylene 6.—A mixture of thymine 18 (0.4 g, 3.2 mmol) and NaH (60%; 0.25 g, 6.3 mmol) in DMSO (20 cm<sup>3</sup>) was stirred for 1 h at room temperature under N<sub>2</sub>. Tetrachloroethylene 6 (1.57 g, 9.5 mmol) was then added and the mixture was stirred for 15 h. TLC in solvent S<sub>7</sub> indicated the presence of title product 20 and a small amount of compound 19. The volatile components were evaporated off and the residue was chromatographed in solvent  $S_7$  to give compound 20 (0.15 g, 20%), m.p. 225-226 °C after crystallization from solvent S7 (Found: C, 38.25; H, 2.8; Cl, 31.9; N, 12.7. C<sub>7</sub>H<sub>6</sub>Cl<sub>2</sub>N<sub>2</sub>O<sub>2</sub> requires C, 38.04; H, 2.74; Cl, 32.08; N, 12.67%);  $\lambda_{max}/nm$  260 ( $\epsilon$  10 300);  $\delta_{H}$  11.75 (1 H, s, NH), 7.60 (1 H, s, 2'-H), 7.31 (1 H, d, 6-H) and 1.77 (3 H, d, Me);  $\delta_{\rm C}$  164.29, 148.58, 138.78, 112.18 and 12.33 (thymine), 127.26 and 120.88 (C-1', -2'); m/z (EI) 224, 222 and 220 (M, 0.6, 3 and 3%), 187, 185 (M - Cl, 37 and 100), 181, 179 and 177 (M - HCNO, 2, 12 and 19), 144 and 142 (M - HCNO - Cl, 8, 25) and 116 and 114 (M - HCNO - CO - Cl, 20, 42).

B. From thymine 18 and dichloroacetylene. Dichloroacetylene-diethyl ether complex <sup>7</sup> (2.5 cm<sup>3</sup>, ~4 mmol) was added to a solution of thymine 18 (0.25 g, 2 mmol) in THF-HMPA (2:1; 30 cm<sup>3</sup>). The mixture was stirred for 27 h at room temperature whereupon it was evaporated (see compound 16, method B). Chromatography in solvent S<sub>8</sub> gave dichloro enamine 20 (53 mg, 12%), m.p. 230-231 °C, which was identical (TLC, UV and <sup>1</sup>H NMR) with the product obtained by method A.

(E)-N<sup>1</sup>-(1,2-Dichlorovinyl)thymine 20 and (E,E)-N<sup>1</sup>,N<sup>3</sup>-Bis-1,2-dichlorovinyl)thymine 21.—A mixture of thymine 18 (0.5 g, 4 mmol) and NaH (60%; 0.32 g, 7.9 mmol) in HMPA (10 cm<sup>3</sup>) was stirred with trichloroethylene 7 (1.55 g, 11.8 mmol) for 15 h at room temperature. The usual work-up and chromatography in solvent S<sub>8</sub> gave compound 21 (0.36 g, 28%), m.p. 155-156 °C after crystallization from solvent S<sub>8</sub> (Found: C, 34.4; H, 2.1; Cl, 45.0; N, 8.8. C<sub>9</sub>H<sub>6</sub>Cl<sub>4</sub>N<sub>2</sub>O<sub>2</sub> requires C, 34.21; H, 1.91; Cl, 44.88; N, 8.87%);  $\lambda_{max}/nm$  264 ( $\epsilon$  10 000);  $\delta_{H}(500$  MHz) 7.915 and 7.911 (1 H, 2 d, 6-H), 7.463, 7.443, 7.409 and 7.383 (2 H, 4 s, 1'- and 2"-H), 1.881 and 1.873 (3 H, 2 d, Me);  $\delta_{\rm C}(125 \text{ MHz})$ 160.425 and 160.263 (C-4), 146.024 and 145.913 (C-2), 139.525 and 139.340 (C-6), 126.371 (C-1'), 123.853 and 123.609 (C-1"), 122.110, 121.995 and 121.829 (C-2', -2"), 111.508 (C-5) and 12.555 (Me); m/z (CI) 321, 319, 317 and 315 (M + H, 11, 48, 100 and 76%), 285, 283, 281 and 279 (M - Cl, 2, 21, 63 and 66), 181, 179 and 177 (M - ClCH=CClN=C=O, 0.8, 6, 9%), 144, 142 (M - ClCH=CCl-N=C=O - Cl, 5 and 16) and 116 and 114 (M - ClCH=CCl-N=C=O - CO - Cl, 8 and 17)

Further elution afforded compound 20 (50 mg, 5.7%) which was identical (TLC, IR and <sup>1</sup>H NMR) with the product obtained from thymine 18 and tetrachloroethylene 6 in DMSO.

N<sup>1</sup>-(*Trichlorovinyl*)*cytosine* **23**.—The procedure for preparation of compound **11** was employed using cytosine **22** (1.11 g, 10 mmol), NaH (57%; 0.84 g, 20 mmol) and tetrachloroethylene **6** (9.94 g, 60 mmol) in HMPA (70 cm<sup>3</sup>). The crude product was chromatographed in solvent system S<sub>2</sub> to give *compound* **23** (0.5 g, 20.8%), m.p. 229–230 °C after crystallization from the same solvent (50 mg/25 cm<sup>3</sup>) (Found: C, 30.2; H, 1.8; Cl, 44.4; N, 17.3. C<sub>6</sub>H<sub>4</sub>Cl<sub>3</sub>N<sub>3</sub>O requires C, 29.97; H, 1.68; Cl, 44.23; N, 17.47%);  $\lambda_{max}/nm 250 \ (\varepsilon 9900)$ ;  $\delta_{\rm H} 7.72 \ (2 \text{ H, s, NH}_2)$ , 7.65 (1 H, d, 6-H) and 5.87 (1 H, d, 5-H);  $\delta_{\rm C}$  166.79, 152.76, 144.31 and 97.18 (cytosine) and 127.91 and 123.60 (C-1', -2'): m/z (CI) 246, 244, 242 and 240 (M + H, 3, 25, 78 and 82%) and 208, 206 and 204 (M - Cl, 3, 65 and 100).

(E)-N<sup>1</sup>-(1,2-*Dichlorovinyl*)*cytosine* **24**.—The procedure for the preparation of compound **16** was followed with cytosine **22** (0.22 g, 2 mmol). The usual work-up and chromatography in solvent system S<sub>2</sub> afforded *compound* **24** (0.1 g, 24.2%), m.p. 219–220 °C after crystallization from solvent system S<sub>3</sub> (Found: C, 34.9; H, 2.4; Cl, 34.6; N, 20.2. C<sub>6</sub>H<sub>5</sub>Cl<sub>2</sub>N<sub>3</sub>O requires C, 34.98; H, 2.45; Cl, 34.42; N, 20.40%);  $\lambda_{max}/nm$  251 ( $\epsilon$  8300);  $\delta_{\rm H}$ 7.56 (2 H, s, NH<sub>2</sub>), 7.53 (1 H, d, 6-H), 7.10 (1 H, s, 2'-H) and 5.81 (1 H, d, 5-H);  $\delta_{\rm C}$  166.81, 152.78, 144.64 and 96.58 (cytosine), 129.94 and 118.47 (C-1', -2'); m/z (CI) 210, 208 and 206 (M + H, 11, 64 and 100%) and 172 and 170 (M - Cl, 12 and 32).

# (E)- $N^{1}$ -(1,2-Dichlorovinyl)- $N^{4}$ -(dimethylaminomethylene)-

cytosine 25.—Compound 24 (65 mg, 0.32 mmol) was stirred with N,N-dimethylformamide dimethyl acetal (0.11 g, 0.95 mmol) in DMF (2 cm<sup>3</sup>) for 15 h at room temperature. The volatile components were removed under reduced pressure (oilpump) and the residue was triturated with hexane to give compound 25 (70 mg, 85%), m.p. 214–215 °C (decomp.) after crystallization from ethyl acetate-acetone (1:1) (Found: C, 41.4; H, 4.0; Cl, 27.1; N, 21.2. C<sub>9</sub>H<sub>10</sub>Cl<sub>2</sub>N<sub>4</sub>O requires C, 41.40; H, 3.86; Cl, 27.16; N, 21.2. C<sub>9</sub>H<sub>10</sub>Cl<sub>2</sub>N<sub>4</sub>O requires C, 41.40; H, 3.86; Cl, 27.16; N, 21.46%);  $\lambda_{max}/nm$  318 ( $\varepsilon$  29 700);  $\delta_{H}$ (CDCl<sub>3</sub>) 8.80 (1 H, s, Me<sub>2</sub>NCH=), 7.16 (1 H, d, 6-H), 6.45 (1 H, s, 2'-H), 6.06 (1 H, d, 5-H) and 3.13 and 3.10 (6 H, 2 s, Me<sub>2</sub>N); m/z (CI) 265, 263 and 261 (M + H, 9, 54 and 82%), and 227 and 225 (M - Cl, 33 and 100).

N<sup>6</sup>,N<sup>6</sup>-Dibenzoyl-N<sup>9</sup>-(trichlorovinyl)adenine 26.—Pyridine was repeatedly evaporated under reduced pressure from compound 11 (0.3 g, 1.1 mmol). Fresh pyridine (15 cm<sup>3</sup>) was added to the residue followed by benzoyl chloride (0.32 g, 2.3 mmol) and the mixture was stirred for 15 h at room temperature. The volatile components were removed under reduced pressure, the last portions by evaporation with toluene (15 cm<sup>3</sup>). Chromatography in solvent system S<sub>8</sub> gave compound 26 (0.3 g, 56%), m.p. 195-196 °C after crystallization from hexane-acetone (1:1) (Found: C, 53.1; H, 2.4; Cl, 22.4; N, 15.05.  $C_{21}H_{12}Cl_3N_5O_2$  requires C, 53.36; H, 2.56; Cl, 22.50; N, 14.82%);  $\lambda_{max}/nm$  248 ( $\varepsilon$  24 400) and 275sh (17 900);  $\delta_{\rm H}({\rm CDCl}_3)$  8.75 and 8.12 (2 H, 2 s, 2- and 8-H) and 7.85 (4 H, d), 7.50 (2 H, t) and 7.37 (4 H, t) (2  $\times$  Ph);  $\delta_{\rm C}$  172.11 (CO), 153.54, 152.67, 152.24, 142.90 and 119.20 (adenine), 133.88, 133.20, 129.44 and 128.78 (Ph), and 126.49 and 126.35 (C-1', -2'); m/z (EI) 475, 473 and 471 (M, 1, 4 and 4%).

(E)-N<sup>9</sup>-(1-Chloro-2-methoxyvinyl)adenine 27.—A mixture of compound 16 (34 mg, 0.15 mmol) and MeONa (25% in MeOH; 0.1 cm<sup>3</sup>, 0.44 mmol) in DMF (3 cm<sup>3</sup>) was stirred for 5.5 h at room temperature whereupon it was evaporated. The residue was washed several times with solvent system  $S_2$  (15 cm<sup>3</sup>). The combined washings were evaporated and the crude product was chromatographed in the same solvent mixture to give compound 27 (21 mg, 63%), m.p. 162-163 °C after crystallization from solvent system S<sub>1</sub> (Found: C, 42.8; H, 3.7; Cl, 15.6; N, 31.0. C<sub>8</sub>H<sub>8</sub>ClN<sub>5</sub>O requires C, 42.59; H, 3.57; Cl, 15.71; N, 31.04%);  $\lambda_{max}/nm 258$  ( $\epsilon 16 000$ );  $\delta_{H} 8.21$  and 8.15 (2 H, 2 s, 2- and 8-H), 7.39 (2 H, s, NH<sub>2</sub>), 7.17 (1 H, s, H-2') and 3.60 (3 H, s, Me);  $\delta_{\rm C}$ 156.15, 153.50, 149.61, 140.91 and 118.04 (adenine), 147.43 (C-2'), 102.33 (C-1') and 61.04 (Me); m/z (EI) 227 and 225 (M, 33 and 100%), 212 and 210 (M - Me, 17 and 56), 190 (M - Cl, 76), 184 and 182 (M - Me - CO, 28 and 89), 160 (63), 157 and 155 (M - HCN - Me - CO, 20 and 65), and 130 and 128

 $(M - 2 \times HCN - Me - CO, 17 \text{ and } 55)$  (Found: M<sup>+</sup>, 225.0414. C<sub>8</sub>H<sub>8</sub><sup>35</sup>ClN<sub>5</sub>O requires M, 225.0417).

N<sup>9</sup>-Ethynyladenine 30.—A solution of compound 11 (0.6 g, 2.3 mmol) in THF (45 cm<sup>3</sup>) was cooled to -70 °C. BuLi in pentane (2 mol dm<sup>-3</sup>; 4.5 cm<sup>3</sup>, 9 mmol) was then added dropwise under N<sub>2</sub> to the stirred mixture during 10 min. The mixture was stirred at -70 °C for 1 h. The reaction was quenched with methanol (4 cm<sup>3</sup>) and 10% aq. NH<sub>4</sub>Cl (9 cm<sup>3</sup>) and the mixture was brought to room temperature. The solvents were removed under reduced pressure and methanol was repeatedly evaporated from the residue which was then chromatographed in solvent system  $S_1$ . The faster moving dichloroenamine 16 (50 mg, 9.5%) was followed by ynamine 30 (206 mg, 57%), m.p. 191-192 °C (decomp.) after crystallization from ethanol (50 mg/50 cm<sup>3</sup>) and drying at room temperature. Drying at higher temperature led to decomposition (polymerization) (Found: C, 53.0; H, 3.2; N, 43.9. C<sub>7</sub>H<sub>5</sub>N<sub>5</sub> requires C, 52.81; H, 3.17; N, 44.02%);  $v_{\text{max}}/\text{cm}^{-1}$  2140 (C=C);  $\lambda_{\text{max}}/\text{nm}$  258 ( $\varepsilon$  11 500);  $\delta_{\text{H}}$  8.48 and 8.20 (2 H, 2 s, 2- and 8-H), 7.57 (2 H, s, NH<sub>2</sub>) and 4.63 (1 H, s, 2'-H); δ<sub>c</sub> 156.34, 154.19, 151.18, 141.49 and 116.75 (adenine), 68.98 (C-1') and 65.85 (C-2'); m/z (EI) 159 (M, 100%), 132 (M - HCN, 24), and 105  $(M - 2 \times HCN, 8)$  (Found: M<sup>+</sup>, 159.0543. C<sub>7</sub>H<sub>5</sub>N<sub>5</sub> requires M, 159.0545).

2,6-Diamino-N<sup>9</sup>-ethynylpurine **31**.—The procedure for ynamine **30** was modified as follows. A solution of compound **12** (1 g, 3.6 mmol) in THF (50 cm<sup>3</sup>) was cooled to -70 °C, 2 mol dm<sup>-3</sup> BuLi in pentane (7.1 cm<sup>3</sup>, 14.2 mmol) was added and the reaction mixture was stirred at -70 °C for 3 h. The usual work-up and chromatography (solvent S<sub>2</sub>) gave a *tan solid* **31** (0.26 g, 42%), m.p. 198–200 °C (decomp.) after crystallization from methanol (Found: C, 48.5; H, 3.5; N, 48.1. C<sub>7</sub>H<sub>6</sub>N<sub>6</sub> requires C, 48.26; H, 3.47; N, 48.27%);  $v_{max}/cm^{-1}$  2140 (C=C);  $\lambda_{max}/nm$  275 ( $\varepsilon$  10 100);  $\delta_{\rm H}$  7.99 (1 H, s, 8-H), 6.96 (2 H, s, 6-NH<sub>2</sub>), 6.17 (2 H, s, 2-NH<sub>2</sub>) and 4.42 (1 H, s, 2'-H);  $\delta_{\rm C}$  161.89, 156.95, 154.21, 137.75 and 111.33 (purine), 70.40 (C-1',  $J_{1',2'-\rm H}$  57.9) and 65.39 (C-2',  $J_{2',2'-\rm H}$  265.5); m/z (EI) 174 (M, 100%), 132 (M – HN=C=NH, 16) (Found: M<sup>+</sup>, 174.0653. C<sub>7</sub>H<sub>6</sub>N<sub>6</sub> requires M, 174.0654).

2-Amino-6-benzyloxy-N<sup>9</sup>-ethynylpurine **32**.—The procedure for ynamine **30** was modified using compound **13** (0.2 g, 0.5 mmol) and 2 mol dm<sup>-3</sup> BuLi (1.1 cm<sup>3</sup>, 2.1 mmol) in THF (15 cm<sup>3</sup>), and a reaction time 2 h at -78 °C. The crude product was chromatographed in solvent system S<sub>7</sub> to give ynamine **32**, which was crystallized twice from hexane–acetone (7:3) (30 mg, 21%), m.p. 200–201 °C (softening from 185 °C) (Found: C, 63.5; H, 4.3; N, 26.2. C<sub>14</sub>H<sub>11</sub>N<sub>5</sub>O requires C, 63.39; H, 4.18; N, 26.40%);  $v_{max}/cm^{-1}$  2140 (C=C);  $\lambda_{max}/nm$  264 ( $\varepsilon$  14 000);  $\delta_{\rm H}(90\%$  pure) 8.16 (1 H, s, 8-H), 7.49 and 7.37 (5 H, m, Ph), 6.89 (2 H, s, NH<sub>2</sub>), 5.46 (2 H, s, CH<sub>2</sub>) and 4.56 (1 H, s, 2'-H); m/z (EI) 265 (M, 45.6%) and 91 (C<sub>6</sub>H<sub>5</sub>CH<sub>2</sub>, 100) (Found: M<sup>+</sup>, 265.0959. C<sub>14</sub>H<sub>11</sub>N<sub>5</sub>O requires M, 265.0964).

N<sup>6</sup>-Benzoyl-N<sup>9</sup>-ethynyladenine 33.—A. From N<sup>6</sup>-benzoyl derivative 14. The procedure for ynamine 30 was followed on a 0.7 mmol scale (THF, 15 cm<sup>3</sup>) at −78 °C. After the usual work-up, compound 33 was isolated by chromatography in solvent system S<sub>4</sub> (85 mg, 47%), m.p. 175–176 °C after crystallization from the same solvent mixture (Found: C, 63.8; H, 3.3; N, 26.4. C<sub>14</sub>H<sub>9</sub>N<sub>5</sub>O requires C, 63.86; H, 3.45; N, 26.61%);  $v_{max}/cm^{-1}$  2150 (C≡C) and 1680 (CO);  $\lambda_{max}/nm$  278 (ε 17 700);  $\delta_{\rm H}$  11.38 (1 H, s, NH), 8.82 and 8.81 (2 H, 2 s, 2- and 8-H), 8.02, 7.63 and 7.53 (5 H, d, t and t, Ph) and 4.79 (1 H, s, 2'-H).

B. From  $N^6$ ,  $N^6$ -dibenzoyl derivative 26. Method A was employed on a 2.1 mmol scale with dibenzoyl derivative 26. TLC (S<sub>4</sub>) showed the presence of four UV-absorbing spots.

Chromatography in solvent system  $S_7$  afforded the major compound 33 (0.1 g, 18%), identical with the product obtained by method A.

N<sup>1</sup>-*Ethynylcytosine* **34**.—The procedure described above was adapted for compound **23** (0.1 g, 0.4 mmol), 2 mol dm<sup>-3</sup> BuLi (0.8 cm<sup>3</sup>, 1.6 mmol) and THF (15 cm<sup>3</sup>), reaction time 2 h. Chromatography in solvent S<sub>2</sub> gave a small amount of dichloro enamine **24** followed by *product* **34** (25 mg, 44%), m.p. > 100 °C (decomp.) (Found: C, 53.25; H, 3.5; N, 30.9. C<sub>6</sub>H<sub>5</sub>N<sub>3</sub>O requires C, 53.32; H, 3.73; N, 31.11%);  $\nu_{max}/cm^{-1}$  2130 (C=C);  $\lambda_{max}/nm$  254 and 282;  $\delta_{H}[(CD_3)_2NCDO, -15 °C]$  7.91 (2 H, s, NH<sub>2</sub>), 7.71 (1 H, d, 6-H), 6.32 (1 H, d, 5-H) and 4.23 (1 H, s, 2'-H);  $\delta_{C}$  165.66, 154.25, 144.84 and 97.40 (cytosine), 75.43 (C-1') and 62.23 (C-2'). The olefinic peaks were present in both <sup>1</sup>H and <sup>13</sup>C NMR spectra.

N<sup>1</sup>-*Ethynylthymine* **35**.—The experiment was performed as in the preceding case. Trichloro enamine **19** (0.4 g, 1.6 mmol), 2 mol dm <sup>3</sup> BuLi (3.1 cm<sup>3</sup>, 6.2 mmol) and THF (25 cm<sup>3</sup>), reaction time 3 h at −70 °C. Chromatography in solvent system S<sub>7</sub> afforded *compound* **35** (0.12 g, 51%), which was crystallized from hexane–acetone (1:1), m.p. 151–152 °C (decomp.) (Found: C, 56.0; H, 4.2; N, 18.45. C<sub>7</sub>H<sub>6</sub>N<sub>2</sub>O<sub>2</sub> requires C, 55.98; H, 4.03; N, 18.67%);  $\nu_{max}$ /cm<sup>-1</sup> 2140 (C≡C);  $\lambda_{max}$ /nm 269 (ε 7200);  $\delta_{\rm H}$  11.67 (1 H, s, NH), 7.62 (1 H, apparent s, 6-H), 4.22 (1 H, s, 2'-H) and 1.73 (3 H, apparent s, Me);  $\delta_{\rm C}$  164.24, 150.71, 139.68, 112.63 and 12.23 (thymine), 73.46 (C-1', J<sub>1',2'-H</sub> 37) and 63.93 (C-2', J<sub>2',2'-H</sub> 245); *m*/z (EI) 150 (M, 34%), 107 (M – HCNO, 80), 79 (M – HCNO – CO, 22), 78 (M – HCNO – CO – H, 34) and 52 (HC≡C–N=CH, 100).

N<sup>9</sup>-*Ethyladenine* **36**.—Ynamine **30** (50 mg, 0.3 mmol) was hydrogenated in ethanol (50 cm<sup>3</sup>) over Pd/C (10%; 60 mg) at 50 psi in a Parr apparatus for 3 h. The solution was filtered (Celite) and the filtrate was evaporated to give the title solid **36** (45 mg, 88%), m.p. 193–194 °C (lit.,<sup>39</sup> 193 °C) identical with an authentic sample.<sup>36</sup>

N<sup>1</sup>-Ethylthymine **37**.—Ynamine **35** (50 mg, 0.33 mmol) was hydrogenated for 5 h as described for compound **36** to give the title solid **37** (44 mg, 86%), m.p. 220–221 °C after crystallization from THF-diethyl ether (7:3) (lit.,<sup>40</sup> 220–222 °C);  $\lambda_{max}/nm$ 270 ( $\varepsilon$  10 300); the <sup>1</sup>H NMR spectrum was similar to that described.<sup>40</sup>

Reaction of Ynamine 30 with Acetone 38.- A solution of ynamine 30 (50 mg, 0.3 mmol) and NaNH<sub>2</sub> (19 mg, 0.5 mmol) in THF  $(30 \text{ cm}^3)$  was stirred at room temperature under N<sub>2</sub> for 25 min. Acetone 38 (70 mm<sup>3</sup>, 0.9 mmol) was then added and the mixture was stirred for an additional 5 h. The progress of the reaction was monitored by TLC in solvent system S<sub>2</sub>. The mixture was cooled to 0-5 °C and the reaction was quenched with methanol (5  $cm^3$ ). The solvents were evaporated off and the crude product was chromatographed using solvent system S<sub>2</sub> as eluent to give carbinol **41** (30 mg, 45%), m.p. 225–230 °C (decomp., softening at 209-211 °C) after crystallization from the same solvent mixture (Found: C, 55.1; H, 5.2; N, 32.1.  $C_{10}H_{11}N_5O$  requires C, 55.29; H, 5.10; N, 32.24%);  $v_{max}/cm^{-1}$ 2260 (C=C);  $\lambda_{max}/nm$  259 ( $\epsilon$  10 200);  $\delta_{H}$  8.40 and 8.20 (2 H, 2 s, 2- and 8-H), 7.50 (2 H, s, NH<sub>2</sub>), 5.62 (1 H, s, OH) and 1.47 (6 H, s, Me);  $\delta_{\rm C}$  156.84, 154.56, 151.42, 141.99 and 117.42 (adenine), 80.36 (COH), 68.12 (C-1'), 64.12 (C-2') and 32.01 (Me); m/z (EI) 217 (M, 50%) and 202 (M – Me, 100) (Found: M<sup>+</sup>, 217.0959. C<sub>10</sub>H<sub>11</sub>N<sub>5</sub>O requires M, 217.0964).

Reaction of Ynamine 30 with Cyclohexanone 39.—The reaction was performed as in the preceding experiment, in THF

(15 cm<sup>3</sup>) with cyclohexanone **39** (61 mg, 0.6 mmol) added. The usual work-up and chromatography in solvent system  $S_2$  afforded *compound* **43** (32 mg, 29%), m.p. 215–217 °C after crystallization from ethanol (Found: C, 64.1; H, 7.2; N, 19.7.  $C_{19}H_{25}N_5O_2$  requires C, 64.19; H, 7.09; N, 19.71%);  $\lambda_{max}/nm$  284sh ( $\varepsilon$  6400), 237 (25 000) and 205 (15 400);  $\delta_H$  8.32 and 8.14 (2 H, 2 s, 2- and 8-H), 7.32 (2 H, s, NH<sub>2</sub>), 6.34 (1 H, s, 5'-H) and 1.58 and 1.38 (20 H, m, cyclohexane);  $\delta_C$  156.49, 153.12, 148.77, 139.31 and 118.19 (adenine), 152.28 (C-6'), 113.95 (C-2'), 91.51 (C-5'), 82.84 (C-4'), 38.47, 37.57, 24.68, 24.52, 23.94 and 22.52 (CH<sub>2</sub>); *m/z* (FAB) 356 (M + H, 36%), 258 (100) and 136 (adenine + H, 49).

Continued elution gave carbinol **42** (56 mg, 70%), m.p. 215–216 °C (Found: C, 60.4; H, 5.8; N, 27.0.  $C_{13}H_{15}N_5O$  requires C, 60.67; H, 5.88; N, 27.23%);  $v_{max}/cm^{-1}$  2260 (C=C);  $\lambda_{max}/nm$  259 ( $\varepsilon$  10 500);  $\delta_{\rm H}$  8.45 and 8.23 (2 H, 2 s, 2- and 8-H), 7.55 (2 H, s, NH<sub>2</sub>), 5.63 (1 H, s, OH) and 1.87, 1.53 and 1.23 (10 H, m, cyclohexane);  $\delta_{\rm C}$  156.30, 154.17, 150.94, 141.38 and 116.90 (adenine), 78.67 (COH), 69.58 (C-1'), 67.03 (C-2') and 24.84 and 22.73 (CH<sub>2</sub>); m/z (EI) 257 (M, 49%), 214 (M – 43, 100) and 135 (adenine, 55) (Found: M<sup>+</sup>, 257.1280.  $C_{13}H_{15}N_5O$  requires M, 257.1277).

Reaction of Ynamine 30 with 1,3-Dibenzyloxyacetone 40.-Sodium amide (12 mg, 0.3 mmol) was added to a solution of ynamine 30 (30 mg, 0.2 mmol) in THF (15 cm<sup>3</sup>) under N<sub>2</sub> at 0-5 °C. The mixture was stirred at room temperature for 10 min whereupon a solution of 1,3-dibenzyloxyacetone<sup>31</sup> 40 (76 mg, 0.28 mmol) in THF (5 cm<sup>3</sup>) was added. The mixture was stirred for 14 h at room temperature. The customary work-up and chromatography in solvent system S<sub>2</sub> gave compound 44 (70 mg, 72% based on 40), m.p. 105-107 °C after crystallization from ethanol (Found: C, 70.2; H, 6.0; N, 10.20. C<sub>41</sub>H<sub>41</sub>N<sub>5</sub>O<sub>6</sub> requires C, 70.36; H, 5.91; N, 10.01%);  $\lambda_{max}/nm$  288sh ( $\varepsilon$  9000), 237 (28 100) and 208 (46 600);  $\delta_{\rm H}$  8.33 and 8.23 (2 H, 2 s, 2- and 8-H), 7.42 (2 H, s, NH<sub>2</sub>), 7.25 (20 H, m, Ph), 6.52 (1 H, s, 5'-H), 4.49 (8 H, s, PhCH<sub>2</sub>) and 3.71 and 3.68 (8 H, 2 s, CH<sub>2</sub>);  $\delta_{\rm C}$ 156.70, 153.56, 148.80, 139.13 and 118.15 (adenine), 146.14 (C-6'), 138.52, 128.81, 128.07 and 127.99 (Ph), 113.91 (C-2'), 92.96 (C-5'), 86.35 (C-4') and 73.30, 72.24 and 71.12 (CH<sub>2</sub>); m/z (FAB) 700 (M + H, 90%) and 176 (100).

Reaction of Ynamine 35 with 1,3-Dibenzyloxyacetone 40.-The procedure for compound 44 was followed with NaNH<sub>2</sub> (25 mg, 0.6 mmol), ynamine 35 (30 mg, 0.2 mmol) and 1,3dibenzyloxyacetone<sup>31</sup> 40 (77 mg, 0.3 mmol) in THF (5 cm<sup>3</sup>). The crude product was chromatographed in solvent system  $S_6$ to give compound 45 (54 mg, 54% based on ketone 40), m.p. 91-92 °C after crystallization from the same solvent mixture (Found: C, 71.25; H, 6.4; N, 3.9; O, 18.7. C<sub>41</sub>H<sub>42</sub>N<sub>2</sub>O<sub>8</sub> requires C, 71.27; H, 6.13; N, 4.06; O, 18.54%);  $\lambda_{max}/nm$  284 ( $\varepsilon$  7100), 232sh (8500) and 208 (33 900);  $\delta_{\rm H}$  11.44 (1 H, s, NH), 7.34 (1 H, d, 6-H), 7.26 (20 H, m, Ph), 5.89 (1 H, s, 5'-H), 4.47 and 4.45 (8 H, 2 s, PhCH<sub>2</sub>), 3.65 and 3.61 (8 H, 2 s, CH<sub>2</sub>) and 1.65 (3 H, d, Me);  $\delta_{\rm C}$  164.12, 150.21, 140.33, 109.40 and 12.66 (thymine), 147.54 (C-6'), 138.55, 138.43, 128.83, 128.15 and 128.03 (Ph), 113.51 (C-2'), 97.30 (C-5'), 86.17 (C-4'), and 73.30, 72.81, 72.28, 72.21 and 71.11 (CH<sub>2</sub>); m/z (FAB) 691 (M + H, 10%) and 87 (100).

N<sup>6</sup>-Dimethylaminomethylene-N<sup>9</sup>-ethynyladenine **46**.—A solution of ynamine **30** (60 mg, 0.4 mmol) and *N*,*N*-dimethylformamide dimethyl acetal (0.13 g, 1.1 mmol) in DMF was stirred overnight at room temperature. The volatile components were evaporated off and the residue was chromatographed in solvent system S<sub>2</sub> to give compound **46** (60 mg, 75%), m.p. 162– 163 °C (softening from 155 °C) (Found: C, 55.9; H, 4.8; N, 39.1. C<sub>10</sub>H<sub>10</sub>N<sub>6</sub> requires C, 56.05; H, 4.71; N, 39.24%);  $v_{max}/cm^{-1}$  1098

2130 (C=C);  $\lambda_{max}/nm$  315 ( $\varepsilon$  25000);  $\delta_{H}$  8.88 (1 H, s, Me<sub>2</sub>NC*H*=), 8.58 and 8.46 (2 H, 2 s, 2- and 8-H), 4.69 (1 H, s, 2'-H) and 3.18 and 3.11 (6 H, 2 s, Me<sub>2</sub>N); m/z (EI) 214 (M, 82%), 199 (M – Me, 79.1), 158 (M – HCN – NMe, 100) and 144 (M – N=CH–NMe<sub>2</sub> + H, 57).

Adenosine Deaminase Assay.—Compound 30 (3 µmol) was dissolved in warm 0.05 mol dm<sup>-3</sup> Na<sub>2</sub>HPO<sub>4</sub> (pH 7.5; 0.2 cm<sup>3</sup>). After cooling of the mixture to room temperature, a solution of adenosine deaminase (Type VIII, Sigma Chemical Co., St. Louis, Missouri, USA, 0.4 unit) in the same buffer (0.4 cm<sup>3</sup>) was added. The incubation was continued for 3 days and the progress of reaction was followed periodically by TLC in solvent system S<sub>2</sub>) and by UV spectroscopy. A large amount (~90%) of compound 30 was deaminated.

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